

#### **Project Title**

Syndromic Respiratory Testing: Evaluation of Biofire FilmArray Pneumonia Panel

#### **Project Lead and Members**

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#### **Organisation(s) Involved**

Tan Tock Seng Hospital

#### Healthcare Family Group(s) Involved in this Project

Allied Health

#### **Applicable Specialty or Discipline**

Laboratory Medicine

#### **Project Period**

Start date: Not Available

Completed date: Not Available

#### Aims

To evaluate the role of the BioFire Pneumonia Panel in syndromic testing of respiratory infections

#### Background

See poster attached/ below

#### Methods

See poster attached/ below



#### Results

See poster attached/ below

#### Lessons Learnt

Not Available

#### Conclusion

See poster attached/ below

#### **Project Category**

Technology

Medtech

Care & Process Redesign

Productivity, Manhour Savings, Quality Improvement, Job Effectiveness

#### Keywords

Respiratory Infection, Microbiology, Specimen Testing, Polymerase Chain Reaction, Pathogens

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# Syndromic respiratory testing Evaluation of Biofire<sup>®</sup> FilmArray<sup>®</sup> pneumonia panel

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### INTRODUCTION

Respiratory infections are a leading cause of mortality. Conventional microbiology methods require multiple different specimens for different testing platforms – blood, urine, and sputum are sent for culture, antigen detection, and PCR (polymerase chain reaction) analysis.

The BioFire<sup>®</sup> Pneumonia panel is a multiplex PCR system that identifies the 33 most common clinically relevant pathogens from a single specimen. We seek to evaluate the role this panel can play in the syndromic testing of respiratory infections.

### METHODOLOGIES

We evaluated 18 sputum, 27 endotracheal aspirates (ETA) and 11 bronchoalveolar lavage (BAL) specimens from patients with pneumonia. The specimens were run concurrently on the BioFire<sup>®</sup> pneumonia panel and the standard of care multi-test approaches including culture, viral and atypical PCRs. Significance of bacterial organisms were determined using quantitative "binning" by BioFire<sup>®</sup> and concordance between routine laboratory testing methods (culture, viral PCR and PCR for atypical pathogens).

### RESULTS

## CONCLUSION

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The BioFire<sup>®</sup> Pneumonia panel complements culture for the identification of pathogens in the diagnosis of respiratory infections. This PCR panel may be leveraged to obtain a clinical diagnosis within hours compared to days or weeks that current diagnostic measures require. Particularly in the intensive care setting, earlier diagnosis will lead to early targeted clinical intervention, improving patient outcomes.

Having a single test platform also reduces manpower required for the processing of multiple tests on different platforms. Consolidating testing delivers value to the patient with greater convenience, timely results, and reduced healthcare costs.

From the 56 specimens evaluated, the BioFire<sup>®</sup> panel identified a total of 77 bacterial pathogens compared to 24 from conventional respiratory cultures. Legionella pneumophila was detected with the multiplex panel and BioFire<sup>®</sup> from one ETA sample. Rhinovirus A/B/C was detected in a sample sent for respiratory virus multiplex PCR, no viruses were picked up by Biofire<sup>®</sup>.

42 of 56 specimens (75%) were concordant with the BioFire<sup>®</sup> panel and current laboratory testing. The highest concordance was observed in 10/11 BAL samples (91%) followed by 22/27 ETA samples (81%) while sputum samples showed the least concordance at 10/18 (56%).

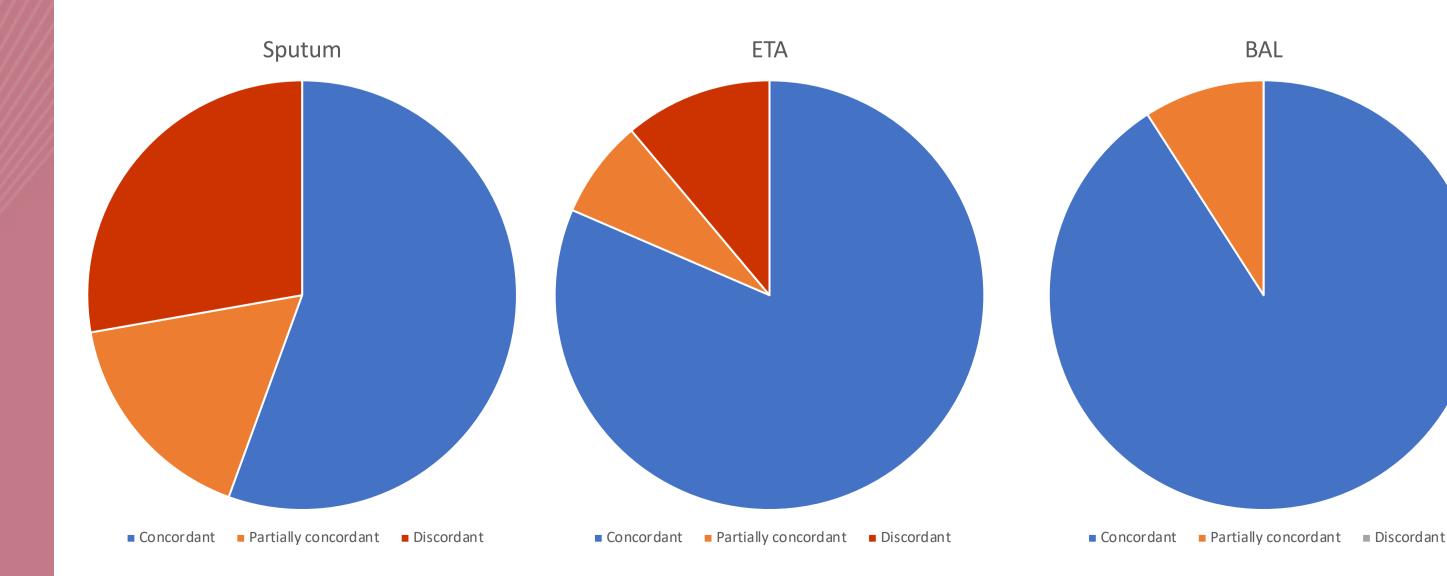
Etiological agents of ventilator-associated pneumonia like Stenotrophomonas maltophilia and Burkholderia *cepacia* are not targeted by BioFire markers.

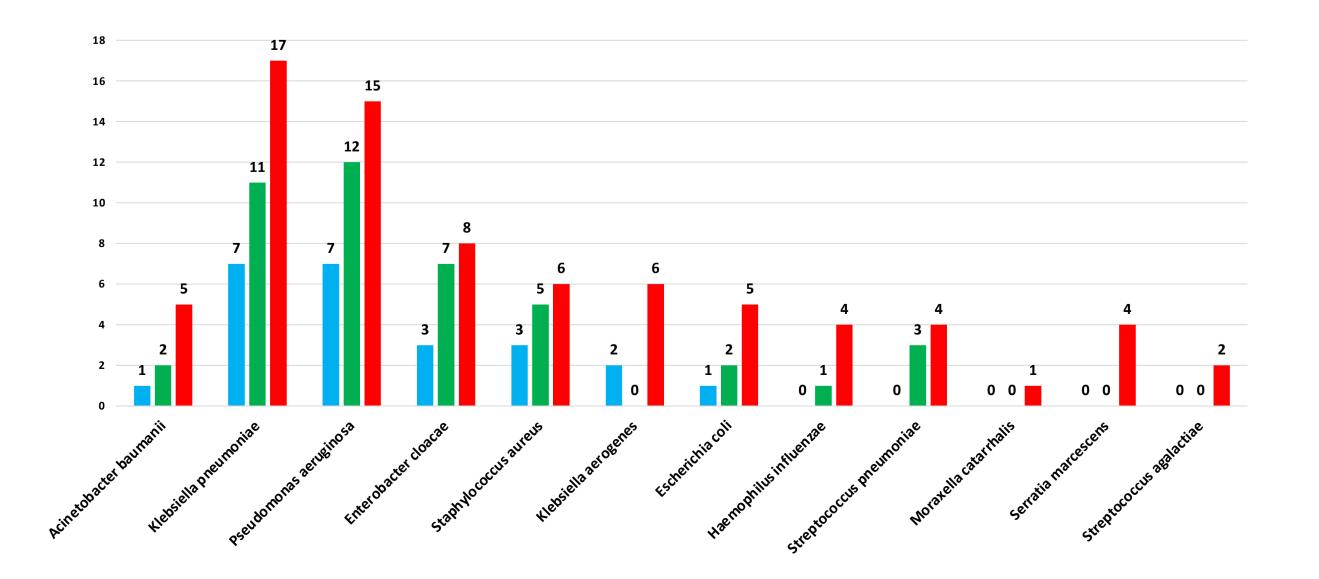
Further studies should include a larger sample size and clinical involvement in order to best evaluate findings.

### FIGURES/ DIAGRAMS

Concordance rates (Sputum, ETA, BAL)

Bacterial detections by BioFire<sup>®</sup> vs culture (Sputum, ETA, BAL)





#### ■ Culture ■ Significant BF isolates ■ Total BF isolates

### Figure 1: Concordance between routine laboratory testing and BioFire® across Sputum, ETA and BAL

Figure 2: Detection rate of each observed bacteria isolates between culture and BioFire®